

# The impacts of habitat change on the genetic diversity of Amsonia tharpii, a rare plant species



## Introduction and Background

- Species are threatened by habitat fragmentation caused by land use change. Habitat fragmentation can lead to population decline.
- Inbreeding, genetic drift, and small population size can decrease genetic diversity. Low genetic diversity lowers species' ability to adapt, increasing extinction risk, as does small population size.
- Amsonia tharpii is found in 5 populations in New Mexico (4) and Texas (1) (Fig. 1a), which are threatened by gas and oil mining. Increased drought frequency due to climate change also imperils A. tharpii.
- To understand the impacts of habitat change on the historical genetic diversity and population structure of *A. tharpii*, it was compared to two other Amsonia species, A. longiflora and A. fugatei.
- Amsonia longiflora differs from the other two species by having a corolla tube that is almost twice as long (Fig. 1b). It also has the largest distribution of the three species, occurring in southern NM, Texas and Mexico.
- Amsonia fugatei is morphologically similar to A. tharpii, endemic to Socorro County, NM, and only known from three small populations (Fig. 1a).
- Results will help to inform the Fish and Wildlife service whether or not A. tharpii should be listed under the Endangered Species Act.

### Objective and Hypothesis

We expect to find that the geographically closest populations of *A. tharpii* will be most closely related. We also expect A. tharpii to have less genetic diversity and higher inbreeding than the widespread A. longiflora. Conversely, we expect A. tharpii to have more genetic diversity and less inbreeding than A. fugatei.

## Methods

- Field Work: Leaf tissue was collected and dried on silica gel in 2020 and 2022. We sampled all three populations of A. fugatei, all five of A. tharpii, and six of A. longiflora. A total of 10 individuals per population were used for genetic analyses.
- **DNA Extraction**: We used a modified CTAB protocol and quantified DNA with a Qubit.
- **Sequencing**: We used double digest Restriction-site associated DNA sequencing (ddRadSeq) to generate SNPs. Genomic libraries were prepared at CBG and sequenced at Northwestern University using a NovaSeq.
- Analyses: Raw sequences were demultiplexed, filtered, and aligned using Stacks. VCF files were filtered for missingness and sequence depth using vcftools. Hierfstat and dartR were used in RStudio to estimate genetic diversity and inbreeding statistics. ADMIXTURE was used to investigate population structure and to determine the best number of genetic clusters using the cross validation method.

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## Madison Dieffenbach<sup>1,2</sup>, Dylan Cohen<sup>3</sup>, Alissa Doucet<sup>1,3</sup>, Krissa Skogen<sup>3</sup> <sup>1</sup>Northwestern University- Evanston, IL <sup>2</sup>Madisonmdd@gmail.com <sup>3</sup>Chicago Botanic Garden- Glencoe,



(A. fugatei, A. longiflora, and A. tharpii) is located. (b) Flowers of A. longiflora (c) Flowers of A. tharpii. (d) Non-flowering A. tharpii plant with a Hwkmoth on the plant .



Table. 1								
Stacks parameters (m-M-n+r)	Loci retained/variant sites/ after filtering/ % missing	Ht	Fis					
2_4_4_r60	394/284/91 (75%)	0.153	0.496					
3_5_5_r60	7177/6029/441 (85%)	0.192	0.695					
2_3_3_r80	44547/38326/1114 (25%)	0.133	0.047					
	Stacks Stacks   parameters (m-M-n+r)   2_4_4_r60 3_5_5_r60   2_3_3_r80 3_0	Stacks parameters (m-M-n+r)   Loci retained/variant sites/ after filtering/ % missing     2_4_4_r60   394/284/91 (75%)     3_5_5_r60   7177/6029/441 (85%)     2_3_3_r80   44547/38326/1114 (25%)	Stacks parameters (m-M-n+r)   Loci retained/variant sites/ after filtering/ % missing   Ht     2_4_4_r60   394/284/91 (75%)   0.153     3_5_5_r60   7177/6029/441 (85%)   0.192     2_3_3_r80   44547/38326/1114 (25%)   0.133					









Fig. 2a

Figure 2. Results from ADMIXTURE for (a) A. longiflora (K = 2-8), (b) A. fugatei (K = 2-5), and (c) A. tharpii (K = 2-6). The best K barplot for each species was determined using the cross-entropy method, and is indicated with an arrow.





Fig. 1c



### Table. 2

opulation	Но	Не	Fis	I	hl
CAP	0.113	0.119	0.050	0.011	0.085
CPC	0.110	0.115	0.047	0.009	0.086
TEX	0.110	0.115	0.041	0.003	0.079
RED	0.113	0.126	0.104	0.067	0.144
BEN	0.120	0.118	-0.010	-0.051	0.028

**Table 2.** Population level statistics for *A. tharpii* (Ho = observed heterozygosity, He = expected heterozygosity, Fis = inbreeding coefficient, II-hl Bootstrap confidence interval for Fis).

### **Genetic Diversity**

- in their genetic diversity (Table 1). A. longiflora and A. fugatei.
- similar to that of *A. tharpii* (Table 1)<sup>3</sup>.

#### **Genetic Structure**

- *fugatei* might be generalist pollinated.

- to CPC than TEX.

#### **Preliminary conclusions**

- No strong population structure. gene flow than A. tharpii.

### **Future Directions**

- contemporary genetic diversity.

- Molecular ecology, 22(11), 3124-3140.
- Naturalist, 17(3), 456-469.

### Discussion

• Amsonia tharpii had the lowest genetic diversity and inbreeding coefficient (Table 1). Inbreeding was highest in A. longiflora and A. fugatei, but this was not reflected

• Poor sequencing results may have inflated values for

• Amsonia ludoviciana has a narrow distribution in the southeastern U.S., and was found to have a genetic diversity (0.1918) and inbreeding coefficient (0.129),

• Pollination biology of *Amsonia* is unknown. The long corolla tubes of *A. longiflora* may be visited by hawkmoths, while the short corollas of A. tharpii and A.

 ADMIXTURE results suggest that hawkmoths may be facilitating pollen transfer for each species.

• Amsonia tharpii populations were split into two clusters, BEN and TEX, and CAP, CPC, and RED, with little gene flow between the two (Fig. 2c). • This refutes our hypothesis that populations close together would be more related, since BEN is closer

• Amsonia tharpii is likely self-incompatible.

• Amsonia fugatei and A. longiflora have high inbreeding.

• A. fugatei and A. longiflora show higher levels of

• Sequence *A. tharpii* offspring to determine • Resequence A. longiflora and A. fugatei for comparison • Reproductive and pollination studies to determine

Amsonia self compatibility and pollinators.

### References

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